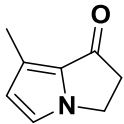
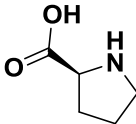
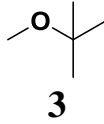
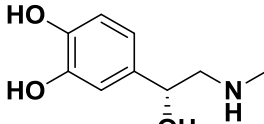
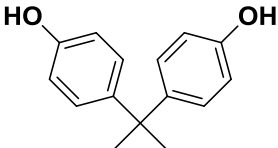
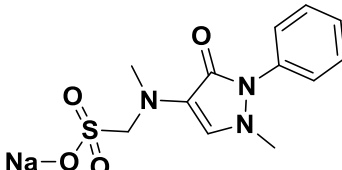
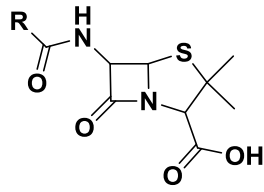
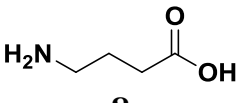
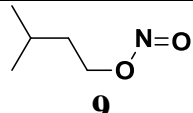
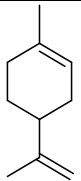
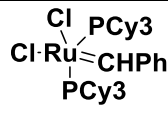

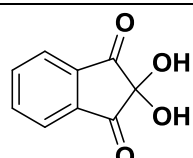
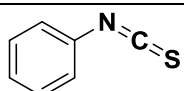
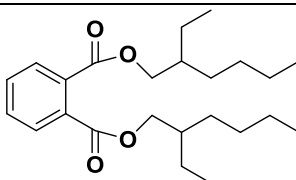
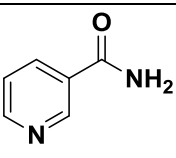
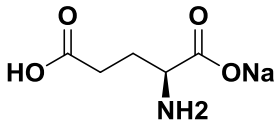
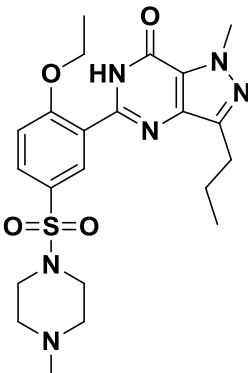
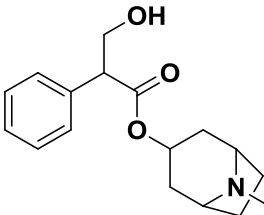
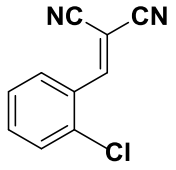


M Широко используемый анальгетик
N Антидетонационная добавка к моторным топливам
O Реагент для секвенирования белков (реагент Эдмана)
P Компонент эпоксидных смол
R Гормон, вырабатываемый в организме в моменты повышенного нервного возбуждения
Q Открытие этого соединения произвело переворот в медицине, автор открытия получил Нобелевскую премию
S Боевое отравляющее вещество
T Сосудорасширяющее средство, используется как противоядие при отравлении цианидами

 1	 2	 3	 4
 5	 6	 7	 8
 9	 10	 11	 12
 13	 14	 15	 16
 17	 18	 19	 20

Результаты оформите в таблицу.

A	B	C	D	E	F	G	H	I	J
K	L	M	N	O	P	R	Q	S	T

Задача №2

При анализе минерального удобрения – сульфат-нитрата аммония, являющегося смесью $(\text{NH}_4)_2\text{SO}_4$ и NH_4NO_3 , в отдельной навеске в 1,560 г вначале было определено общее содержание азота. Для этого весь азот из обеих солей был выделен в виде аммиака (действием щелочи, а для NO_3^- , кроме того, и действием восстановителя – металлического алюминия). Выделенный аммиак отогнали в 50,00 мл 0,5250 моль-экв/л раствора H_2SO_4 , а затем оставшийся избыток H_2SO_4 оттитровали 6,40 мл 0,3750 моль-экв/л раствора NaOH .

Другую навеску 1,370 г кипятили с 50,00 мл того же 0,3750 моль-экв/л раствора NaOH ; при этом вытесняется NH_3 только из NH_4^+ обеих солей. После отгонки NH_3 избыток NaOH оттитровывали 7,14 мл 0,5250 моль-экв/л раствора H_2SO_4 .

Напишите все описанные реакции. Рассчитайте процентное содержание NH_4NO_3 и $(\text{NH}_4)_2\text{SO}_4$ в удобрении.

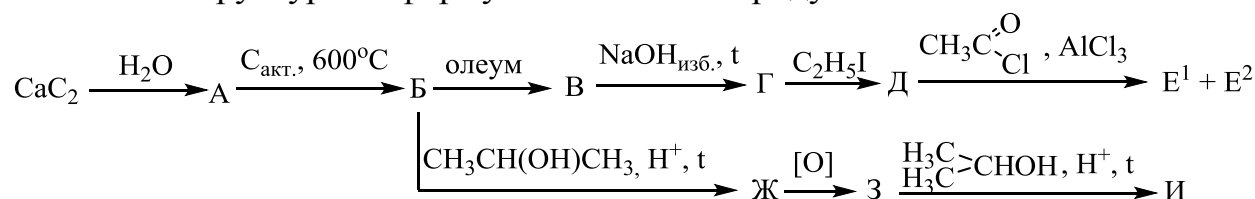
Задача №3

Найденные при археологических раскопках останки человека были подвергнуты анализу на содержание радиоактивного изотопа углерода ^{14}C . Образцы показали удельную активность живых d , равную $10.3 \text{ распадов} \cdot \text{мин}^{-1} \cdot \text{г}^{-1}$. Известно, что удельная активность живых организмов d_0 равна $15.3 \text{ распадов} \cdot \text{мин}^{-1} \cdot \text{г}^{-1}$, а период полураспада углерода ^{14}C , $t_{1/2}$, равен 5580 лет.

Определите возраст этих останков. Оцените неточность радиоуглеродного метода оценки возраста в археологических исследованиях, считая, что период полураспада определен с высокой степенью точности, а число распадов с высокой степенью точности, а число распадов с ошибкой в 1%.

Задача №4

Напишите структурные формулы и назовите продукты А-И:



Задача №5

Одним из наиболее активных нестероидных противовоспалительных препаратов является индометацин. Он находит применение при лечении ревмокардита, инфекционного полиартрита, остеоартрита, болезни Бехтерева и других заболеваний, сопровождающихся воспалением.

Впервые индометацин был получен Т. Шеном [Т. Y. Shen, T. B. Windholz, A. Rosegay, B. E. Witzel, et. al. Non-Steroid Antiinflammatory agents. // J. Am. Chem. Soc. 1963. — Vol. 85, N. 4. -P. 488–489].

Заполните схему: напишите ключевые продукты в синтезе индометацина.

The reaction scheme shows the synthesis of indomethacin starting from 4-methoxyaniline and 4-oxopentanoic acid tert-butyl ester. The starting material 4-methoxyaniline is a benzene ring with a methoxy group (MeO) at the top and an amino group (NH₂) at the bottom. The second starting material is 4-oxopentanoic acid tert-butyl ester, shown as a five-carbon chain with a ketone group (C=O) at the second carbon and a tert-butyl ester group (CO₂Bu-t) at the fourth carbon. The reaction proceeds through four steps:

- 4-methoxyaniline reacts with 1. HCl/NaNO₂ and 2. Sn/HCl to form product **A**.
- Product **A** reacts with 4-oxopentanoic acid tert-butyl ester to form product **B**.
- Product **B** reacts with HCl/EtOH to form product **C**.
- Product **C** reacts with 1. NaOH(aq)/ pClC₆H₄COCl and 2. 210 °C to form product **D**.

4

selective precipitation,¹¹ or modification of the antigen.^{8,12}

An approach to the isolation of antibody against a soluble protein antigen was suggested by Campbell, Leuscher, and Lerman.¹³ They coupled a soluble protein antigen to an insoluble, modified (*p*-aminobenzyl-) cellulose by means of a diazonium bond. This provided an insoluble protein antigen which could specifically combine with antibody and upon acidification to pH 3.2 be separated by centrifugation into soluble antibody and insoluble antigen. This method provided a yield of 57% of the total antibody present in the original serum as determined by quantitative precipitation with specific antigen. The purity (precipitable antibody/total protein) of the soluble material recovered by specific adsorption and subsequent elution was 90%.

This problem of immune adsorbents was re-investigated in our Laboratory as a result of commercially available *p*-aminobenzylcellulose, Cellex-PAB, (Bio-Rad Lab., Richmond, Calif.) and the coupling of a simple haptenic material, arsanilic acid, to cellulose (ICN Corp., City of Industry, Calif.). The method employed in our study involves the use of an antigen "fixed" by a diazo bond to the commercially available cellulose to which the *p*-aminobenzyl group has been coupled. After exposure of the appropriate antiserum to the antigen-coupled Cellex, the column was washed prior to elution of antibody with a glycine-HCl buffer pH 3.0. Utilizing this method we have isolated rabbit antibodies to (1) bovine serum albumin (BSA), (2) whole ragweed pollen extracts (WRE), (3) a highly purified Timothy pollen fraction¹⁴ and (4) a simple hapten (arsanilate).

In order to couple the protein antigens to the Cellex-PAB diazotization was carried out at 0° with 2 *N* HCl and 14% NaNO₂ for 1 hour. The diazotized Cellex-PAB was filtered and washed successively with 5% sodium acetate, 5% urea, and distilled water. Coupling of the antigen was accomplished by addition of the diazotized Cellex to the protein solution at pH 8, and the suspension was magnetically stirred overnight at 2°. The non-reacting antigen was recovered by filtration, and the Cellex was further coupled with β -naphthol to block any unreacted free diazonium sites. The antigen-coupled Cellex was washed successively with distilled water, glycine-HCl buffer, pH 3.0, and was then readjusted to pH 7.25 with 0.1 *N* NaOH. The immune adsorbent then was poured as a slurry into a 1 × 10 cm. column, packed by gravity at 2°, and washed with an appropriate buffer (pH 7.25).

The isolation of purified rabbit anti-whole ragweed pollen extract (AWRE) antibodies from a whole ragweed-coupled Cellex (WRE) column was accomplished in the following manner. To 1 g. samples of WRE coupled-Cellex was added 20 ml. of rabbit immune serum containing 50 mg. of antibody protein precipitable by WRE. The time required for the serum to pass through the column was 1-2 hours. The column was then washed with 50-100 ml. of a citrate-phosphate buffer, pH 7.25, in order to remove all non-specific proteins. The specifically bound antibody then was dissociated from the immune adsorbent with a glycine-HCl buffer, pH 3.0. Five 5-ml. fractions were collected, and adjusted to pH 7.4 with 0.1 *N* NaOH. The presence of antibody in the collected fractions was

determined by ring tests and a quantitative precipitin test. The recovery of protein from the eluted fractions varied from 24-35 mg., which represented a yield of 46-70%, and 83% of the recovered protein was precipitable with specific antigen.

The same general procedure was used in the isolation of rabbit antibodies directed against bovine serum albumin, arsanilate hapten, and a highly purified Timothy pollen fraction. It also has been applied to the isolation of human reaginic antibody from non-treated Timothy sensitive patients,¹⁵ and these results will be presented in more detail in a subsequent publication.

This work was supported by a grant from the U. S. Public Health Service; Grant No. AI-01339.

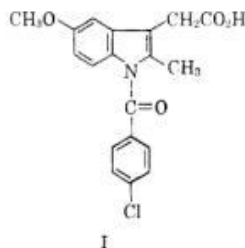
(15) A. Malley, *Federation Proc.*, **21**, No. 2, 14 (1962).

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PASADENA, CALIFORNIA DAN H. CAMPBELL
RECEIVED DECEMBER 19, 1962

NON-STEROID ANTI-INFLAMMATORY AGENTS

Sir:

We wish to report a new class of anti-inflammatory and antipyretic agents, substituted indole acetic and propionic acids. Of some three hundred and fifty indole derivatives studied, one member of the series, 1-(*p*-chlorobenzoyl)-5-methoxy-2-methylindole-3-acetic acid (I), designated as indomethacin, has demonstrated a high degree of anti-inflammatory activity both in the granuloma inhibition assay¹ and in the foot-edema



assay.² Indomethacin was equally active in intact or adrenalectomized animals and was also active by local application directly to the cotton pellets. Its anti-inflammatory potency relative to phenylbutazone varied from ten to eighty-five times depending on the test employed.³ Fever induced in rabbits by intravenous injection of pyrogenic lipopolysaccharide prepared from *E. coli* was effectively blocked by I subcutaneously administered. Antipyretic potency of I was approximately 10 times that of aminopyrine and 20 times phenylbutazone, with duration of action resembling the latter compound. In animals indomethacin is relatively free of activities referable to central nervous, autonomic or cardiovascular systems.

For the synthesis of indomethacin, 5-methoxy-2-methylindole-3-acetic acid⁴ was converted to its anhydride with dicyclohexylcarbodiimide in tetrahydrofuran. The anhydride was treated with zinc chloride and *t*-butanol to give *t*-butyl 5-methoxy-2-methyl-3-indolylacetate, m.p. 110-111°. Acylation of the *t*-butyl ester with *p*-chlorobenzoyl chloride afforded *t*-butyl 1-(*p*-

(1) R. Meier, W. Schuler and P. Desaulles, *Experientia*, **6**, 469 (1950), as modified by C. A. Winter and C. C. Porter, *J. Am. Pharm. Assoc., Sci. Ed.*, **46**, 515 (1957).

(2) C. A. Winter, E. A. Risley and G. W. Nuss, forthcoming publications.

(3) Preliminary clinical reports of indomethacin indicate that its potency in man is about 0.25-0.5 that of cortisone.

(4) E. Shaw, *J. Am. Chem. Soc.*, **77**, 4319 (1955).

(11) S. I. Epstein, P. Doty and W. C. Boyd, *J. Am. Chem. Soc.*, **78**, 3306 (1956).

(12) S. J. Singer, J. E. Fothergill and J. R. Shainoff, *ibid.*, **81**, 2277 (1959).

(13) D. H. Campbell, E. Leuscher and L. S. Lerman, *Proc. Natl. Acad. Sci. U. S.*, **37**, 575 (1951).

(14) A. Malley, A. Lietze and C. E. Reed, *J. Allergy*, **33**, 84 (1962).

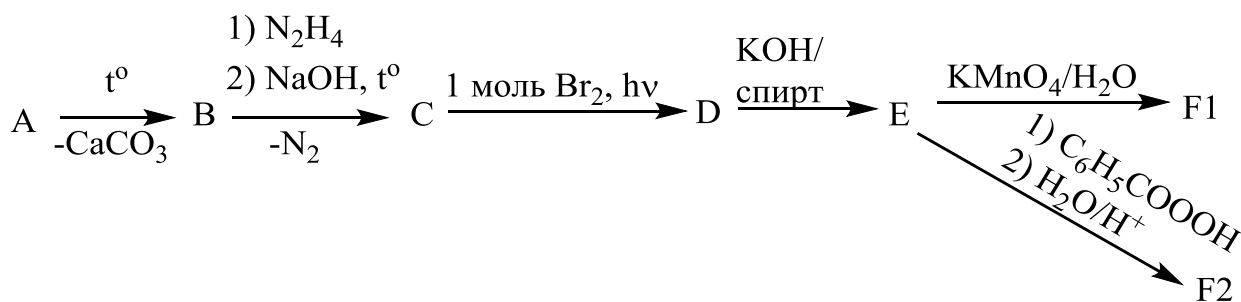
Задача №6

Некоторая реакция протекает при температуре 20°С за 24 ч, а при 40°С – за 4 ч. Оцените время протекания реакции при температуре 60°С, полагая, что время реакции обратно пропорционально константе скорости. Сформулируйте правило

Вант-Гоффа, рассчитайте температурный коэффициент и подтвердите выполнимость правила для данной реакции в исследуемом диапазоне температур (20-60 °С).

Задача №7

Напишите структурные формулы соединений А-Ф, зная, что вещество А является кальциевой солью пимелиновой кислоты ($\text{HOOC}(\text{CH}_2)_5\text{COOH}$):



1. Укажите различия в структуре соединений F1 и F2.
2. Напишите структурную формулу вещества G, из которого может быть получена пимелиновая кислота, если известно, что оно образуется при взаимодействии этилена с четыреххлористым углеродом CCl_4 в соотношении 3:1 в присутствии перекиси.

Задача №8

0.727 г D-рибозы ($\text{C}_5\text{H}_{10}\text{O}_5$) поместили в калориметр и сожгли в избытке кислорода. Подъем температуры составил 0.910 К. При сгорании в тех же условиях 0.825 г бензойной кислоты температура возросла на 1.940 К.

Рассчитайте изменение внутренней энергии при сгорании D-рибозы и энтальпию ее образования, если изменение внутренней энергии при сгорании бензойной кислоты равно ($-3251 \text{ кДж}\cdot\text{моль}^{-1}$).